## **Bridging Converts a Noncytotoxic nor-Paclitaxel Derivative to a Cytotoxic Analogue by Constraining It to the T-Taxol Conformation**

**Shoubin Tang,† Chao Yang,† Peggy Brodie,† Susan Bane,‡ Rudravajhala Ravindra,‡ Shubhada Sharma,‡ Yi Jiang,§ James P. Snyder,§ and David G. I. Kingston\*,†**

*Department of Chemistry, M/C 0212, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061, Department of Chemistry, State University of New York at Binghamton, Binghamton, New York 13902, and Department of Chemistry, Emory Uni*V*ersity, Atlanta, Georgia 30322*

*dkingston@*V*t.edu*

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**The synthesis of the bridged A-nor-paclitaxel 4 has been achieved from paclitaxel in a key test of the T-Taxol conformational hypothesis. Although the unbridged A-nor-paclitaxel 3 is essentially noncytotoxic, the bridged analogue 4 is strongly cytotoxic. This result provides strong evidence for the T-Taxol conformation as the bioactive tubulin-binding conformation of paclitaxel.**

The natural product paclitaxel (PTX) (Taxol, **1**) is a clinically approved drug for several tumor malignancies, $<sup>1</sup>$  and its</sup> chemistry and biology have been investigated extensively.2 It acts by promoting the polymerization of tubulin to stabilized microtubules, leading to apoptotic cell death, $3-6$ 

and its clinical activity is believed to be directly related to this microtubule-binding activity.6



Since the bioactivity of PTX is intimately connected with its tubulin-assembly properties and since tubulin assembly is initiated by the noncovalent binding of paclitaxel to tubulin, the nature of this binding is a matter of great interest.

*Sci. U.S.A.* **<sup>1993</sup>**, *<sup>90</sup>*, 9552-9556. (6) Blagosklonny, M. V.; Fojo, T. *Int. J. Cancer* **<sup>1999</sup>**, *<sup>83</sup>*, 151-156.

<sup>†</sup> Virginia Polytechnic Institute and State University.

<sup>‡</sup> State University of New York at Binghamton.

<sup>§</sup> Emory University.

<sup>(1) (</sup>a) Rowinsky, E. K. *Annu. Re*V*. Med.* **<sup>1997</sup>**, *<sup>48</sup>*, 353-374. (b) Crown, J.; O'Leary, M. *Lancet* **<sup>2000</sup>**, *<sup>355</sup>*, 1176-1178.

<sup>(2) (</sup>a) Kingston, D. G. I. 2005 In *Anticancer Agents from Natural Products*; Cragg, G. M., Kingston, D. G. I., Newman, D. J. Eds.; CRC Press: Boca Raton, 2005; pp 89-122. (b) Ojima, I.; Kuduk, S.; Chakravarty, <sup>S</sup>*. Ad*V*. Med. Chem.* **<sup>1999</sup>**, *<sup>4</sup>*, 69-124.

<sup>(3)</sup> Schiff, P. B.; Fant, J.; Horwitz, S. B. *Nature* **<sup>1979</sup>**, *<sup>277</sup>*, 665-667. (4) Horwitz, S. B. *Trends Pharmacol. Sci.* **<sup>1992</sup>**, *<sup>13</sup>*, 134-136.

<sup>(5)</sup> Jordan, M. A.; Toso, R. J.; Thrower, D.; Wilson, L. *Proc. Natl. Acad.*

In one scenario, a knowledge of the tubulin-binding conformation of paclitaxel would enable the design of simple nontaxoid compounds with comparable binding affinity and bioactivity.

Although the taxane ring system of paclitaxel is relatively rigid, the compound has flexible side chains at C2, C4, C10, and notably C13. As a result, there are many possible conformations available for binding to tubulin. Two different paclitaxel models of the tubulin-binding conformation were proposed on the basis of NMR observables and molecular modeling. A "nonpolar" conformation was put forth on the basis of solution NMR investigations in nonpolar solvents,  $7-9$ whereas similar studies in polar solvents were interpreted to favor a bound "polar" conformation.<sup>10-13</sup> Although most of the reports assumed a single conformation, deconvolution of PTX in  $CDCl<sub>3</sub><sup>14</sup>$  and  $D<sub>2</sub>O/DMSO-d<sub>6</sub><sup>15</sup>$  makes it clear that the molecule adopts  $9-10$  conformations, no one of which achieves a population above 30%.

A second approach focused on tubulin-bound paclitaxel in the solid state. Application of REDOR NMR provided  $F^{-13}$ C distances of 9.8 and 10.3 Å between the fluorine of a 2-(*p*-fluorobenzoyl)PTX and the C3′ amide carbonyl and C3′ methine carbons, respectively.16 A related examination reported a distance of 6.5 Å between the fluorines of 2-(*p*fluorobenzoyl)-3′-(*p*-fluorophenyl)-10-acetyldocetaxel and, like the first solid state study, proposed the polar form to be tubulin-bound.17-<sup>19</sup>

The "polar" and "nonpolar" conformations have inspired several elegant synthetic studies designed to generate constrained analogues that maintain these conformations, but none of these constrained analogues have shown tubulinpolymerization or cytotoxic activities equal to or greater than those of PTX itself. Various compounds designed to mimic the "polar" conformation were either inactive<sup>20</sup> or less active than PTX.21 Analogues based on the "nonpolar" conformation were also less active than PTX.22-<sup>25</sup>

- (7) Dubois, J.; Guenard, D.; Gueritte-Voeglein, F.; Guedira, N.; Potier, P.; Gillet, B.; Betoeil, J.-C. *Tetrahedron* **<sup>1993</sup>**, *<sup>49</sup>*, 6533-6544.
- (8) Williams, H. J.; Scott, A. I.; Dieden, R. A.; Swindell, C. S.; Chirlian, L. E.; Francl, M. M.; Heerding, J. M.; Krauss, N. E. *Can. J. Chem.* **1994**, *<sup>72</sup>*, 252-260.
- (9) Cachau, R. E.; Gussio, R.; Beutler, J. A.; Chmurny, G. N.; Hilton, B. D.; Muschik, G. M.; Erickson, J. W. *Supercomput. Appl. High Perform. Comput.* **<sup>1994</sup>**, *<sup>8</sup>*, 24-34.
- $(10)$  Vander Velde, D. G.; Georg, G. I.; Grunewald, G. L.; Gunn, C. W.; Mitscher, L. A. J. Am. Chem. Soc. 1993,  $115$ ,  $11650-11651$ .
- W.; Mitscher, L. A. *J. Am. Chem. Soc.* **<sup>1993</sup>**, *<sup>115</sup>*, 11650-11651. (11) Paloma, L. G.; Guy, R. K.; Wrasidlo, W.; Nicolaou, K. C. *Chem. Biol.* **<sup>1994</sup>**, *<sup>1</sup>*, 107-112.
- (12) Ojima, I.; Chakravarty, S.; Inoue, T.; Lin, S.; He, L.; Horwitz, S. B.; Kuduk, S. C.; Danishefsky, S. J. *Proc. Natl. Acad. Sci. U. S.A*. **1999**,
- *<sup>96</sup>*, 4256-4261.
- (13) Ojima, I.; Kuduk, S. D.; Chakravarty, S.; Ourevitch, M.; Begue, J.-P. *J. Am. Chem. Soc.* **<sup>1997</sup>**, *<sup>119</sup>*, 5519-5527.
- (14) Snyder, J. P.; Nevins, N.; Cicero, D. O.; Jansen, J. *J. Am. Chem. Soc.* **<sup>2000</sup>**, *<sup>122</sup>*, 724-725.
- (15) Snyder, J. P.; Nevins, N.; Jimenez-Barbero, Cicero, D.; Jansen, J. M. Unpublished work.
- (16) Li, Y.; Poliks, B.; Cegelski, L.; Poliks, M.; Gryczynski, Z.; Piszczek, G.; Jagtap, P. G.; Studelska, D. R.; Kingston, D. G. I.; Schaefer, J.; Bane,
- S. *Biochemistry* **<sup>2000</sup>**, *<sup>39</sup>*, 281-291. (17) Ojima, I.; Inoue, T.; Chakravarty, S. *J. Fluorine Chem.* **1999**, *97*,
- $3 10$ .

(18) The X-ray crystal structure of polar PTX (ref 19) utilized in ref 16 shows an F $\cdots$ F distance of 4.8 Å when the *para*-hydrogens of the phenyl<br>rings at the C-2 and C-3' positions are replaced with fluorines  $(r_{C-F})$ rings at the C-2 and C-3<sup>'</sup> positions are replaced with fluorines ( $r_{\text{(C-F)}}$  = 1.33 Å) 1.33 Å).

A more fruitful approach stems from the electron crystallographic (EC) structure of  $\alpha\beta$ -tubulin stabilized by  $\text{Zn}^{2+}$ and PTX. It provides a detailed view of subunit structure but at 3.7 Å resolution leaves the PTX conformation unresolved.26 A model derived from intersection of PTX NMR and X-ray conformations with the EC density led to the proposal of T-Taxol as the bound conformation.27 A concurrent EC refinement<sup>28</sup> at  $3.5$  Å resolution provided additional confidence in the model. Following these developments, a T-like conformer with a reorganized C-13 side chain (PTX-NY) was proposed as an alternative binding model.<sup>29,30</sup> However, this conformer is inconsistent with the EC density.<sup>31,32</sup>

We report here an indirect method to establish the nature of the tubulin-taxane binding conformation more firmly. As described in more detail elsewhere,  $33$  we previously designed and prepared several bridged analogues such as **2** based on the T-Taxol or butterfly conformation. The latter juxtaposes the C3′-phenyl and C4-OAc groups and encouraged the construction of taxanes with bridges linking these positions. Three of the analogues have tubulin-assembly and cytotoxic activities superior to those of paclitaxel, providing strong support for the T-Taxol conformation.<sup>33,34</sup> Nonetheless, the actual improvement in bioactivity is relatively modest, varying from a 22-fold increase in cytotoxicity in the A2780 bioassay to a factor of 1.4 in the case of the PC3 prostate cancer cell line.<sup>33</sup>



- (19) Mastropaolo, D.; Camerman, A.; Luo, Y.; Brayer, G. D.; Camerman, N. *Proc. Natl. Acad. Sci. U.S.A.* **<sup>1995</sup>**, *<sup>92</sup>*, 6920-6924.
- (20) Boge, T. C.; Wu, Z.-J.; Himes, R. H.; Vander Velde, D. G.; Georg, G. I. *Bioorg. Med. Chem. Lett.* **<sup>1999</sup>**, *<sup>9</sup>*, 3047-3052.
- (21) Ojima, I.; Lin, S.; Inoue, T.; Miller, M. L.; Borella, C. P.; Geng, X.; Walsh, J. J. *J. Am. Chem. Soc.* **<sup>2000</sup>**, *<sup>122</sup>*, 5343-5353.
- (22) Ojima, I.; Geng, X.; Lin, S.; Pera, P.; Bernacki, R. J. *Bioorg. Med. Chem. Lett.* **<sup>2002</sup>**, *<sup>12</sup>*, 349-352.
- (23) Geng, X.; Miller, M. L.; Lin, S.; Ojima, I. *Org. Lett.* **<sup>2003</sup>**, *<sup>5</sup>*, 3733- 3736.
- (24) Querolle, O.; Dubois, J.; Thoret, S.; Dupont, C.; Guéritte, F.; Guénard, D. *Eur. J. Org. Chem.* 2003, 542-550.
- (25) Querolle, O.; Dubois, J.; Thoret, S.; Roussi, F.; Montiel-Smith, S.; Guéritte, F.; Guénard, D. J. Med. Chem. 2003, 46, 3623-3630.
- (26) Nogales, E.; Wolf, S. G; Downing, K. H. *Nature* **<sup>1998</sup>**, *<sup>391</sup>*, 199- 203.
- (27) Snyder, J. P.; Nettles, J. H.; Cornett, B.; Downing, K. H.; Nogales, E. *Proc. Natl. Acad. Sci. U.S.A.* **<sup>2001</sup>**, *<sup>98</sup>*, 5312-5316.
- (28) Lowe, J.; Li, H.; Downing, K. H.; Nogales, E. *J. Mol. Biol.* **2001**, *<sup>313</sup>*, 1045-1057.
- (29) Geney, R.; Sun, L.; Pera, P.; Bernacki, R. J.; Xia, S.; Horwitz, S. B.; Simmerling, C. L.; Ojima, I. *Chem. Biol.* **<sup>2005</sup>**, *<sup>12</sup>*, 339-348.
- (30) The T-Taxol conformer proposed by Geney, Ojima, and colleagues was named "REDOR-Taxol".<sup>29</sup> However, since there are upwards of 100 PTX conformations that meet the REDOR constraints including T-Taxol, 31,32
- we refer to the Geney-Ojima model as New York paclitaxel (PTX-NY). (31) Johnson, S. A.; Alcaraz, A.; Snyder, J. P. *Org. Lett.* **<sup>2005</sup>**, *<sup>7</sup>*, 5549- 5552.
- (32) Alcaraz, A. A.; Mehta, A. K.; Johnson, S. A.; Snyder, J. P. *J. Med. Chem*. **<sup>2006</sup>**, *<sup>49</sup>*, 2478-2488.

Tubulin-assembly activity appears to be less sensitive to structural variations than cytotoxicity, but  $2$  had an  $ED_{50}$  for tubulin polymerization of  $0.21 \mu M$  as compared with paclitaxel's  $0.50 \mu M^{33}$  We thus elected to carry out a rigorous test of the T-Taxol hypothesis by determining the effect of bridging on the bioactivity of the much less active starting compound A-*nor*-paclitaxel (**3**).

A-*nor*-Paclitaxel, or  $(1\alpha)$ -15(16)-anhydro-11(15<sup>---</sup>1)-abeotaxol (**3**), is an A-ring contracted paclitaxel analogue that was first reported by us in 1991.<sup>35</sup> Biological studies revealed that **3** was much less cytotoxic than paclitaxel toward the KB cell line, but that it retained ability to inhibit tubulin disassembly at a level about one-third that of paclitaxel.36 A preliminary molecular modeling study showed that the rearranged A-*nor*-baccatin core of A*-nor-*paclitaxel has a conformation similar to that of the baccatin core of paclitaxel. As a result, a number of A-*nor*-paclitaxel analogues with modifications on the C-1 isopropenyl moiety and the C-2 benzoyl group were prepared. A few of these analogues showed enhanced tubulin binding activities, in some cases to the same level as that of paclitaxel, but none possessed comparable cytotoxicity.<sup>36,37</sup> Interestingly, unlike PTX, where some modifications on the C-2 benzoyl group increase tubulin-assembly activity,38 the same modifications on the C-2 benzoyl group of A-*nor*-paclitaxel uniformly decrease tubulin-assembly activity. $37$  Based on this information, the bridged compound **4** provides an excellent critical test of the T-Taxol hypothesis, since any cytotoxicity observed would represent a significant increase in bioactivity due to the imposed conformational lock.

We investigated two separate approaches to the synthesis of **4**. The first approach involved the synthesis of the A-*nor*baccatin III **5** and subsequent attachment of a modified side chain and olefin metathesis to form the bridge. This approach was selected because of its flexibility, since it could in principle be adapted to prepare several compounds with different chain lengths.



(33) Ganesh, T.; Guza R. C.; Bane, S.; Ravindra, R.; Shanker, N.; Lakdawala, A. S.; Snyder, J. P.; Kingston, D. G. I. *Proc. Natl. Acad. Sci. U.S.A.* **<sup>2004</sup>**, *<sup>101</sup>*, 10006-10011.

The approach failed, however, at the point of the conversion of the 10-deacetyl derivative **6** to **5**. We thus elected to pursue the simpler direct route of conversion of a preformed bridged derivative to its A-*nor* analogue.

The known compound dihydrobritaxel-5 was prepared as previously described<sup>33</sup> and converted to the silyl-protected compound **7**. This was then treated with SOCl<sub>2</sub> at  $-20$  °C to give the A-*nor* derivative **8** (Scheme 1). The resulting



compound was then deprotected to yield the desired product **4**. The unbridged A-*nor*-paclitaxel **3** was also prepared as previously described<sup>35</sup> for comparison purposes.

PTX and compounds **3** and **4** were evaluated for tubulinassembly capacity, tubulin-binding ability, and cytotoxicity against the A2780 and PC3 cell lines (Table 1). Impressively,

## **Table 1.** Bioactivity of Paclitaxel Analogs



*<sup>a</sup>* GDP-tubulin, determined as described in Supporting Information. *<sup>b</sup>* GMPcPP microtubules, determined as described in Supporting Information. *<sup>c</sup>* Data from ref 33. *<sup>d</sup>* Not determined. *<sup>e</sup>* GTP-tubulin; the value for PTX under these conditions is 0.37  $\mu$ M<sup>3</sup>

the bridged A-*nor*-paclitaxel **4** binds to GMPcPP microtubules with affinity slightly greater than that of PTX itself and is more efficacious by a factor of 2 in lowering the critical concentration of tubulin-GDP. The enhanced tubulin polymerization activity  $(ED_{50})$  of compound 4 is probably due to an increase in both the ligand's efficacy and its affinity for tubulin (Table 1).

The unbridged A-*nor*-analog **3** was only weakly active in all of the assays performed. For example, no inhibition of fluorescent PTX binding to GMPcPP microtubules was

<sup>(34)</sup> Metaferia, B. B.; Hoch, J.; Glass, T. E.; Bane, S. L.; Chatterjee, S. K.; Snyder, J. P.; Lakdawala, A.; Cornett, B.; Kingston, D. G. I. *Org. Lett.* **<sup>2001</sup>**, *<sup>3</sup>*, 2461-2464.

<sup>(35)</sup> Samaranayake, G.; Magri, N. F.; Jitrangsri, C.; Kingston, D. G. I. *J. Org. Chem.* **<sup>1991</sup>**, *<sup>56</sup>*, 5114-5119.

<sup>(36)</sup> Yuan, H.; Kingston, D. G. I.; Long, B. H.; Fairchild, C. A.; Johnston, K. A. *Tetrahedron* **<sup>1999</sup>**, *<sup>55</sup>*, 9089-9100.

<sup>(37)</sup> Chordia, M. D.; Kingston, D. G. I.; Hamel, E.; Lin, C. M.; Long, B. H.; Fairchild, C. A.; Johnston, K. A.; Rose, W. C. *Bioorg. Med. Chem.* **<sup>1997</sup>**, *<sup>5</sup>*, 941-947.

<sup>(38)</sup> Kingston, D. G. I.; Chaudhary, A. G.; Chordia, M. D.; Gharpure, M.; Gunatilaka, A. A. L.; Higgs, P. I.; Rimoldi, J. M.; Samala, L.; Jagtap, P. G.; Giannakakou, P.; Jiang, Y. Q.; Lin, C. M.; Hamel, E.; Long, B. H.; Fairchild, C. R.; Johnston, K. A. *J. Med. Chem.* **<sup>1998</sup>**, *<sup>41</sup>*, 3715-3726.

observed at concentrations of  $3 \text{ up to } 40 \mu\text{M}$  (curves shown in Supporting Information). The association constant for the binding of **3** to microtubules was therefore estimated to be at least 2 orders of magnitude less that that of PTX. The more active tubulin-GTP rather than tubulin-GDP was required to measure critical concentration. The critical concentration of tubulin-GTP in the presence of 1:1 molar ratio of **3** was  $>$  3  $\mu$ M, which is at least 8 times the value determined for PTX under these conditions  $(0.37 \mu M)^3$ 

In cytotoxicity assays, the analogue **4** shows significant cytotoxic activity to the A2780 and the PC3 cell lines, although it is still less active than PTX; in the PC3 cell line it is 3-fold less cytotoxic than PTX (**1**). In contrast, the unbridged A-*nor*-paclitaxel **3** is essentially noncytotoxic, with  $IC_{50}$  values too weak to be determined.

These results indicate that a bridged A-*nor*-paclitaxel, which can maintain a "T-Taxol" conformation, also retains *all of* paclitaxel's tubulin-assembly activity and much of its cytotoxicity. This work thus offers further evidence for the significance of the T-Taxol conformation for tubulin binding and tubulin assembly.

In an attempt to understand the origin of the differences between A-*nor* PTX **3** and bridged A-*nor* PTX **4**, we performed extensive conformational searches for these structures and located 464 and 613 fully optimized conformations, respectively. The latter were examined for the presence of the polar, nonpolar, PTX-NY, and T-Taxol conformers. Bridged **4** delivered 61 T-form minima, while acyclic **3** yielded 75 such conformers. In each of the latter cases, one of the top two fits provides a very close match to the structure of T-Taxol (Figure 1). By comparison, neither the polar nor the PTX-NY types are present in the conformers



**Figure 1.** Superposition of the centroids of the phenyl rings of fully optimized T-forms of the A-*nor* PTX analogues and those of the corresponding rings of T-Taxol (yellow): (a) acyclic **3** (cyan) (rms 0.35 Å); (b) bridged **4** (blue) (rms 0.31 Å).

of **4**, although the acyclic set (**3**) contains them. In addition, the nonpolar form of **4** docks poorly into the binding site (see Supporting Information). Accordingly, we give the latter three rotamers no further consideration.

To compare the compounds in the taxoid binding site, both **3** and **4** were docked into  $\beta$ -tubulin with the Glide protocol using flexible ligand docking and the OPLS force field. Both were found to readily adopt the T-form (see Supporting Information). The MMFF/GBSA $(H_2O)$  energy differences between the docked structures and the respective conformational search global minima for the A-*nor* analogues were computed to show that the energy gap (∆∆*E*) is 1.7 kcal/ mol smaller for bridged **4** than acyclic **3**. With the AMSOL/  $SM5.4PDA/CM1$  aqueous solvation model,<sup>39</sup> we estimated that the free energies of solvation for the two docked structures favor the bridged A-*nor*-Taxol by 0.6 kcal/mol, reducing ∆∆*E* to 1.1 kcal/mol.

To place the latter combination energy difference in the context of the biological data of Table 1, we note that bridged **4** is at least 23- and 600-fold more active than **3** in the A2780 and PC3 cytotoxicity assays, respectively (2000/89 and 8300/ 13.8). If we assume the  $IC_{50}$ 's can be roughly equated with the *K*<sub>i</sub>'s,<sup>40</sup> then  $\Delta \Delta G = -RT \ln[K_i(4)/K_i(3)]$ at 298 K is 1.8 and 3.8 kcal/mol, respectively. These numbers are in reasonable agreement with the calculated value of 1.1 kcal/ mol. They suggest that the source of the activity of **4** versus **3** resides in the relative strain energy associated with binding to tubulin in the T-Taxol conformation offset by a small desolvation penalty.

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**Supporting Information Available:** Detailed synthetic procedures for the synthesis of the bridged A-*nor*-paclitaxel **4** and A-*nor*-baccatin III **5**; <sup>1</sup> H and 13C NMR spectra of compound **4**; tubulin-GTP polymerization by compound **3**; tubulin binding curves for **1**, **3**, and **4**; and conformational searching, conformer extraction and binding site docking for **3** and **4**. This material is available free of charge via the Internet at http://pubs.acs.org.

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<sup>(39)</sup> Hawkins, G. D.; Cramer, C. J.; Truhlar, D. G. *J. Phys. Chem.* **1996**, *100*, 19824-19839.<br>(40) (a) Cheng Y

<sup>(40) (</sup>a) Cheng, Y.-C.; Prusoff, W. H. *Biochem. Pharm.* **<sup>1973</sup>**, *<sup>22</sup>*, 3099- 3108. (b) Cheng, H. C. *J. Pharm. Tox. Methods* **<sup>2002</sup>**, *<sup>46</sup>*, 61-71.